

# Prevalence of antibiotic-resistant bacterial colonization in neonatal intensive care in Europe

Matilda Berkell<sup>1</sup>, Liesbet Van Heirstraeten<sup>1</sup>, Aislinn Cook<sup>2</sup>, Julia Bielicki<sup>3,2</sup>, Surbhi Malhotra-Kumar<sup>1</sup>

<sup>1</sup>Laboratory of Medical Microbiology, Vaccine and Infectious Diseases Institute, University of Antwerp, Antwerp, Belgium,

<sup>2</sup>Centre for Neonatal and Paediatric Infection, St. George's University of London, London, United Kingdom,

<sup>3</sup>Paediatric Research Centre, University of Basel Children's Hospital, Basel, Switzerland

## Introduction

Infants colonized with antibiotic-resistant bacteria (ARB) are at **high-risk** of developing **severe bacterial infections**, that can ultimately lead to sepsis, particularly when born at <32 weeks gestation (high-risk). However, overall **prevalence and burden** of specific antibiotic resistance genes (ARGs) in a high-income setting is largely **unknown**.

## Methodology

In the pan-European NeoIPC project, **skin swabs** ( $N=929$ ) and **stool samples** ( $N=754$ ) were collected during **four point-prevalence surveys** (PPSs) interspaced with 4-, 7-, or 14-day intervals from all infants (<1 year,  $N=468$ ) present in 14 NICUs across six European countries. DNA was extracted (NucliSENS easyMAG, bioMérieux) followed by **RT-qPCR detection** of **carbapenemases** (CBPs), **extended-spectrum-beta-lactamases** (ESBLs), and **vancomycin-resistant enterococci** (VREs) in stools, as well as **methicillin-resistant *Staphylococcus aureus*** (MRSA) in skin swabs (Table 1). Collected anonymized data were analyzed per sample per PPS.

### Abbreviations

**ARB:** antibiotic-resistant bacteria

**ARG:** antibiotic resistance gene

**PPS:** point-prevalence survey

**CBP:** carbapenemase

**ESBL:** extended-spectrum beta-lactamase

**VRE:** vancomycin-resistant enterococci

**MRSA:** methicillin-resistant *Staphylococcus aureus*

**MRCoNS:** methicillin-resistant coagulase-negative staphylococci

**Table 1.** Overview of ARBs/ARGs detected, assays used, and sample matrix utilized for analysis.

Sample matrix	ARB Type	Gene target	Assay	
Stool	CBP	<i>bla</i> <sub>KPC</sub> <i>bla</i> <sub>NDM</sub> <i>bla</i> <sub>VIM</sub> <i>bla</i> <sub>IMP</sub> <i>bla</i> <sub>OXA-48</sub>	Carbaplex-IVD PCR (Bruker)	
		ESBL	<i>bla</i> <sub>CTX-M</sub> group1 <i>bla</i> <sub>CTX-M</sub> group9	Ba04646149_s1, Ba04646127_s1 (ThermoFisher)
			VRE	<i>vanA</i> <i>vanB</i>
Skin swab	MRSA	<i>mecA/mecC</i> <i>SCCmec/orfX</i>	Diarella MRSA-SeqC (Gerbion)	
	MRCoNS	<i>mecA/mecC</i>		

## Results

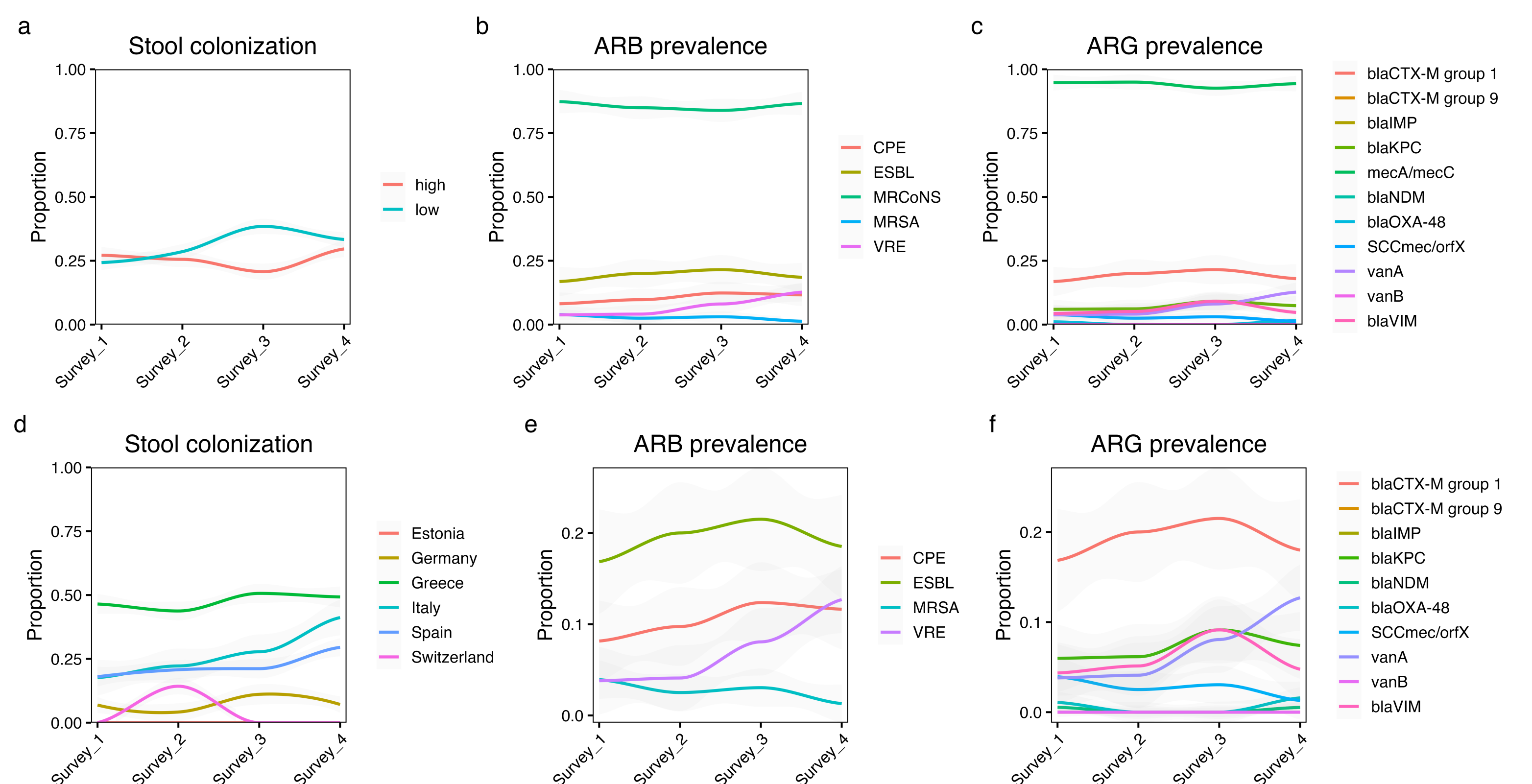
**Table 2.** Antibiotic-resistant bacterial (ARB) colonization patterns across four point-prevalence surveys (PPSs). ARB/ARG colonization rates were determined per PPS by performing PCRs detecting ESBLs, CBPs, and VREs (stool screening), as well as MRSA and MRCoNS (skin swab screening).

Variable	Total samples	Total	Survey-1	Survey-2	Survey-3	Survey-4
Stool colonization	754	217 (29%)	47 (26%)	53 (27%)	57 (31%)	60 (32%)
Number of positive gene targets <sup>1</sup>	754					
1		151 (20%)	36 (20%)	40 (21%)	35 (19%)	40 (21%)
2		48 (6.4%)	9 (4.9%)	10 (5.1%)	14 (7.5%)	15 (7.9%)
3		15 (2.0%)	2 (1.1%)	3 (1.5%)	6 (3.2%)	4 (2.1%)
4		3 (0.4%)	0 (0%)	0 (0%)	2 (1.1%)	1 (0.5%)
VRE detected	754	54 (7.2%)	7 (3.8%)	8 (4.1%)	15 (8.1%)	24 (13%)
<i>vanA</i>		54 (7.2%)	7 (3.8%)	8 (4.1%)	15 (8.1%)	24 (13%)
<i>vanB</i>		0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
CBP detected	754	79 (10%)	15 (8.2%)	19 (9.7%)	23 (12%)	22 (12%)
<i>bla</i> <sub>KPC</sub>		54 (7.2%)	11 (6.0%)	12 (6.2%)	17 (9.1%)	14 (7.4%)
<i>bla</i> <sub>NDM</sub>		2 (0.3%)	1 (0.5%)	0 (0%)	0 (0%)	1 (0.5%)
<i>bla</i> <sub>VIM</sub>		44 (5.8%)	8 (4.3%)	10 (5.1%)	17 (9.1%)	9 (4.8%)
<i>bla</i> <sub>IMP</sub>		0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
<i>bla</i> <sub>OXA-48</sub>		5 (0.7%)	2 (1.1%)	0 (0%)	0 (0%)	3 (1.6%)
ESBL detected	754	145 (19%)	31 (17%)	39 (20%)	40 (22%)	35 (19%)
<i>bla</i> <sub>CTX-M</sub> group1		144 (19%)	31 (17%)	39 (20%)	40 (22%)	34 (18%)
<i>bla</i> <sub>CTX-M</sub> group9		1 (0.1%)	0 (0%)	0 (0%)	0 (0%)	1 (0.5%)
MRSA detected <sup>2</sup>	929	25 (2.7%)	9 (3.9%)	6 (2.5%)	7 (3.0%)	3 (1.3%)
MRCoNS detected <sup>3</sup>	929	821 (88%)	209 (91%)	209 (87%)	200 (87%)	203 (88%)
Risk group <sup>1</sup>	754					
High risk		86 (40%)	22 (47%)	23 (43%)	17 (30%)	24 (40%)
Non-high risk		131 (60%)	25 (53%)	30 (57%)	40 (70%)	36 (60%)
Country <sup>1</sup>	754					
Estonia (N=20)		0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
ES1 (N=20)		0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
Germany		7 (7.1%)	2 (6.9%)	1 (4.2%)	2 (11.1%)	2 (7.1%)
GE1 (N=99)		7 (7.1%)	2 (6.9%)	1 (4.2%)	2 (11.1%)	2 (7.1%)
Greece (N=295)		140 (48%)	33 (46.5%)	35 (43.8%)	39 (50.6%)	33 (49.3%)
GR1 (N=27)		14 (51.9%)	3 (60%)	2 (25%)	7 (88%)	2 (33%)
GR2 (N=69)		54 (78.3%)	11 (65%)	15 (79%)	17 (94%)	11 (73%)
GR3 (N=82)		51 (62.2%)	15 (71%)	13 (59%)	10 (48%)	13 (72%)
GR4 (N=83)		11 (13.3%)	3 (14%)	3 (14%)	2 (9.5%)	3 (16%)
GR5 (N=34)		10 (29.4%)	1 (14%)	2 (22%)	3 (33%)	4 (44%)
Italy (N=70)		19 (27.1%)	3 (17.6%)	4 (22.2%)	5 (27.8%)	7 (41.2%)
IT1 (N=43)		7 (16.3%)	2 (17%)	2 (22%)	0 (0%)	3 (27%)
IT2 (N=27)		12 (44.4%)	1 (20%)	2 (22%)	5 (71%)	4 (67%)
Spain (N=216)		49 (22.7%)	9 (18.0%)	11 (20.8%)	11 (21.2%)	18 (29.5%)
SP1 (N=54)		16 (29.6%)	4 (31%)	3 (25%)	4 (36%)	5 (28%)
SP2 (N=66)		7 (10.6%)	2 (14%)	0 (0%)	0 (0.0%)	5 (29%)
SP3 (N=96)		26 (27.1%)	3 (13%)	8 (36%)	7 (28%)	8 (31%)
Switzerland (N=54)		2 (3.7%)	0 (0%)	2 (14.3%)	0 (0%)	0 (0%)
SW1 (N=44)		2 (4.5%)	0 (0%)	2 (15%)	0 (0%)	0 (0%)
SW2 (N=10)		0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)

<sup>1</sup>Calculated based on stool ARG colonization

<sup>2</sup>MRSA-positive skin swab samples tested positive for both *mecA/mecC* and *SCCmec/orfX*

<sup>3</sup>MRCoNS-positive skin swab samples tested positive only for *mecA/mecC*



**Figure 1.** ARB colonization patterns over time in infants admitted to 14 European NICUs. Panels (a) and (d) display proportion of ARG-positive (CPE, ESBL, or VRE) stool samples defined by infant risk status and country of collection, respectively. Panels (b) and (e) display proportion of ARG/ARB-positive samples with or without MRCoNS, respectively. Finally, panels (c) and (f) display proportion of samples where individual ARGs were detected with and without *mecA/mecC*, respectively.

Overall, **26%** (217/754) stool samples were **ARG-positive** (ESBL, CBP, or VRE) across four PPSs, and in **8.8%** (66/754)  $\geq 2$  ARGs were detected (Table 2). **ESBLs** (19%, *bla*<sub>CTX-M</sub> group1) were **most prevalent**, followed by **CBPs** (10%; mainly *bla*<sub>KPC</sub> and *bla*<sub>VIM</sub>), and **VREs** (7.2%, *vanA*). In contrast, **MRSA** skin colonization was **rare** (2.7%). While not specifically screened for, methicillin-resistant coagulase-negative staphylococci (**MRCoNS**) were detected by the MRSA assay in **majority** of the skin swabs (88%, 821/929).

40% of high-risk infants were colonized, compared to 60% of non-high-risk infants (Pearson's  $\chi^2$ ,  $p=0.3$ ), and **high-risk infants** were more frequently **ESBL-colonized** (21% vs. 16%,  $p=0.086$ ). Gut ARG colonization **varied by country and site** ( $p<0.001$ ), ranging from 0.0%-94% for individual sites, and was most common in Greece (Figure 1).

Colonization rate **stability** over time was found to be **ARG/ARB-dependent**; only **VRE** colonization **increased** from 3.8%-13% during the study period ( $p=0.002$ ), while the **CBP, MRSA, ESBL, and MRCoNS** positivity remained **stable** over time ( $p>0.05$ ).

## Conclusion

- We observed remarkable variation in ARG prevalence across countries and NICU sites.
- Stability of colonization rate/PPS was found to vary for different ARGs.
- Although non-significant, gut colonization with ESBLs, specifically *bla*<sub>CTX-M</sub> group 1, was more frequent in high-risk infants.



University of Antwerp  
Laboratory of Medical Microbiology  
Vaccine and Infectious Disease Institute

Contact: Prof. Surbhi Malhotra-Kumar: [surbhi.malhotra@uantwerpen.be](mailto:surbhi.malhotra@uantwerpen.be)  
Prof. Julia Bielicki: [juliaanna.bielicki@ukbb.ch](mailto:juliaanna.bielicki@ukbb.ch)  
Dr. Matilda Berkell: [matilda.berkell@uantwerpen.be](mailto:matilda.berkell@uantwerpen.be)

Learn more about the NeoIPC project here!

